Original Article

A Novel Methodology for Human Plasma Protein Binding: Prediction, Validation, and Applicability Domain

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ABSTRACT

Background: Plasma protein binding is a key component in drug therapy as it affects the pharmacokinetics and pharmacodynamics of drugs.

Objectives: This study aimed to predict the fraction of plasma protein binding.

Methods: A quantitative structure-activity relationship, convolutional neural network, and feed-forward neural network (QSAR-CNN-FFNN) methodology was used. CNN was used for feature selection, which is known as a difficult task in QSAR studies. The values of the descriptors acquired without the preprocessing procedures were rearranged into matrices, and features from a deep fully connected layer of a pre-trained CNN (ALEXNET) were extracted. Then, the latest features learned from the CNN layers were flattened out and passed through an FFNN to make predictions.

Results: The external accuracy of the validation set (Q^2=0.945, RMSE=0.085) showed the performance of this methodology. Another extremely favorable circumstance of this method is that it does not take a lot of time (only a few minutes) compared to the QSAR-Wrapper-FFNN method (days of hard work and concentration) and it automatically gives us the characteristics that are the best representations of our input.

Conclusion: We can say that this model can be used to predict the fraction of human plasma protein binding for drugs that have not been tested to avoid chemical synthesis and reduce expansive laboratory tests.

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Introduction

Building a quantitative structure-activity relationship (QSAR) model, known as in silico model, is an important step in drug discovery. It allows us to avoid chemical synthesis and reduce expansive and tedious laboratory tests [1, 2]. Different QSAR approaches have been developed over the past few years [3-5]. These approaches can determine the quantitative relations between the variation in the values of calculated descriptors and the biological activity of a series of chemical molecules [6]. Thus, once a correlation has been established, it can be used to predict the property or biological activity of new structures [7, 8]. In QSAR studies, selecting a few relevant descriptors from a large number of features is a difficult task. Feature selection techniques are applied to reduce the number of attributes in the dataset by choosing features that will give us better accuracy with fewer data [9-11]. It also reduces the overfitting and the overtraining risk [12]. Feature selection methods can be divided into three categories: filter, wrapper, and embedded methods. Filters are applied independently of the mapping method used. They are executed before the mapping, to reduce the number of descriptors, following some objective criteria [13]. The wrapper techniques are used to select the optimum subset of features based on the error reduction of classifier algorithms. Wrapper methods perform better than filter methods but are more expensive and time-consuming [14]. Features selected with embedded or hybrid methods are sensitive to the structure of the underlying classifiers. Thus, in most cases, the features selected by one embedded method might not be suitable for others [15].

Plasma protein binding plays a key role in drug therapy that affects the pharmacokinetics and pharmacodynamics of the drug, as they are often directly related to the free drug concentration in plasma [16, 17]. In recent years, several QSAR-artificial intelligence models have been developed to predict plasma protein binding, such as support vector machines and their derivatives [18-20], random forest [21], neural networks [22, 23], and gradient-boosting decision trees [24]. In 2017, Sun et al. constructed QSAR models by six machine-learning algorithms with 26 molecular descriptors [25]. Kumar et al. in 2018 presented a systematic approach using a support vector machine, artificial neural network, K-nearest neighbor, probabilistic neural network, partial least square, and linear discriminant analysis to a diverse dataset of 735 drugs [26]. Yuan et al. in 2020 published a global QSAR model for plasma protein binding and developed a novel strategy to construct a robust QSAR model for plasma protein binding prediction [27]. Recently, deep learning algorithms have attracted the attention of scientists and become an important option for pharmaceutical research. Ramsundar et al. presented deep learning techniques for healthcare that efficiently predict drug activity and structure [28]. Wallach et al. introduced AtomNet, known as the first structure-based deep convolutional neural network (CNN), to predict the bioactivity of small molecules for drug discovery applications [29].

In this work, a novel combined methodology based on QSAR, CNN, and a feed-forward neural network (FFNN) was used to predict plasma protein binding for 277 molecules. The CNN, known to be the most popular algorithm for deep learning, was used for feature selection as an alternative to the wrapper method. The FFNN was then used for the prediction of the plasma protein binding from the extracted features.

Material and Metods

In order to predict the plasma protein binding, a methodology based on five steps was used as summarized in Figure 1: (1) data set collection, (2) molecular descriptors generation, (3) selection of relevant descriptors by a wrapper method and a CNN method, (4) FFNN modeling, and (5) validation of models.

Data set collection

The experimental data of protein binding of the 277 drugs used in this study were selected from the pharmacological basis of the therapeutics handbook [30] and the handbook of clinical drug data [31]. Chemical names and experimental protein binding values are presented in Supplemental Material 1.

Molecular descriptors generation

The numerical representation of molecular structure was assessed in terms of molecular descriptors. The SMILES script (simplified molecular-input line-entry system) required to calculate descriptors was extracted from the open-access database PubChem [32]. SMILES is a specification in the form of a line notation describing the structure of chemical species [33]. The SMILES scripts for the 277 drugs were used to generate 1666 descriptors divided into 20 categories: *(i)* constitutional descriptors, *(ii)* topological descriptors, *(iii)* walk and path counts; *(iv)* connectivity indices; *(v)* index information; *(vi)* 2Dautocorrelations; *(vii)* edge adjacency indices; *(viii)* burden eigenvalue

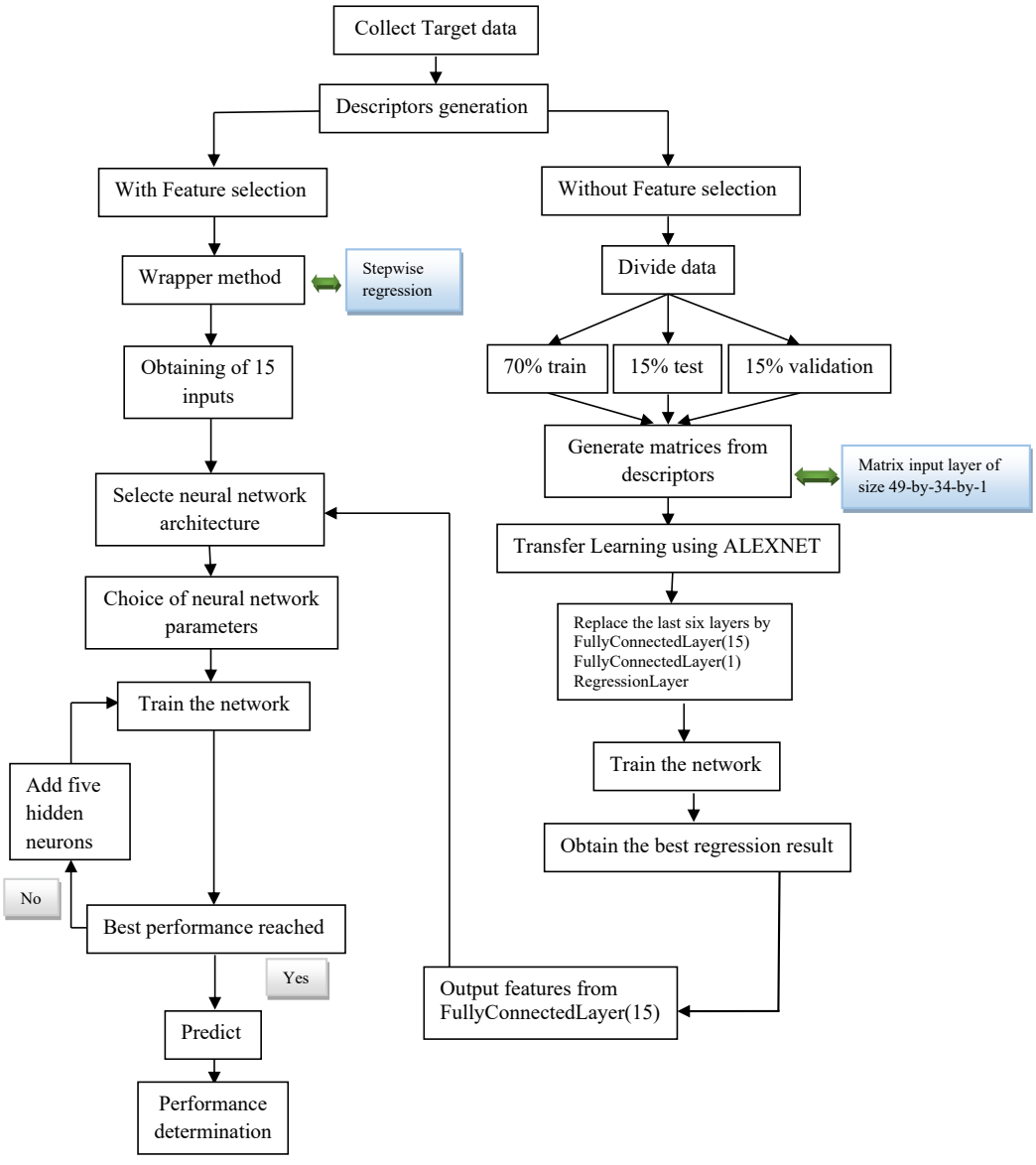


Figure 1. Flow sheet of the procedure followed

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descriptors; *(ix)* topological charge indices; *(x)* eigenvalue-based indices; *(xi)* Randic molecular profiles; *(xii)* geometrical descriptors; *(xiii)* RDF descriptors; *(xiv)* 3D-MoRSE descriptors; *(xv)* WHIM descriptors; *(xvi)* GETAWAY descriptors; *(xvii)* functional group counts; *(xviii)* atom-centered fragments; *(xix)* charge descriptors; and *(xx)* molecular properties.

All descriptors were obtained through the E-Dragon online programs [34], which is known as the electronic remote version of the well-known software DRAGON developed by the Milano Chemometrics and QSAR Research Group by Prof. R. Todeschini. The name and number of calculated descriptors are reported in Supplemental Material 2.

Selection of relevant descriptors

The selection of the most efficient descriptors, which is an important step in QSAR modeling, was made by two techniques in this work.

Wrapper method

The number of molecular descriptors was reduced by the following procedure [35]:

1) Descriptors having constant values (min=max) were eliminated.

2) Quasi-constant descriptors (1st quartile 25%=2nd quartile 75%) were removed.

3) Descriptors with standard relative deviation *RSD*<0.05 were deleted.

The three steps above were performed using STATISTICA software [36].

4) Matrices of the pairwise linear correlation between each pair of columns in the input matrices were calculated via MATLAB [37]. In addition, all variables with a correlation coefficient R>0.75 were removed. For more robustness of the model, the variance inflation factor *VIF* was calculated (Equation 1):

1. $VIF_i=1/(1-R_i^2)$

, Where is the squared correlation coefficient of the i^{th} descriptor versus the remaining ones. All the descriptors with VIF>5 were eliminated from the model [38].

5) The remaining descriptors were analyzed by stepwise regression in STATISTICA software [36]. Only 15 descriptors were selected and used as input variables in the FFNN models.

CNN method

CNNs are one of the most popular algorithms for deep learning, and they are mostly used for image classification. Computers do not see an image as an image, but as an array of pixels and the whole process in CNNs depends on it. Therefore, we provided our CNN network with previously digitalized data as input. Because for deep learning, in image classification, we do not have to understand which features are the best representations of the object, they are taught to us.

In our approach, the 1666 descriptors calculated with E-DRAGON were transformed for each drug into matrices of size 49-by-34-by-1 dimension in MATLAB because CNN requires matrices as inputs [39]. The latest features learned from the CNN layers were flattened out and passed through an FFNN to make predictions.

FFNN modeling

QSAR-wrapper-FFNN

The selected descriptors by the wrapper technique were set as input in an FFNN. So far, there is no clear answer to specify the number of hidden neurons required for a modeling task and it remains an open question. Different approaches are discovering this number, explained in detail in reviews, including methods of selecting the number of hidden nodes in the artificial neural networks review [40]. To decide the number of neurons in the hidden layer, the procedure summarized below was used [41]:

1) In the beginning, only a few hidden neurons (five) were used.

2) The network was trained until the mean square error no longer seemed to improve.

3) At this moment, a few (five) neurons were added to the hidden layer, each with randomly initialized weights, and resumed training.

4) The previous two steps were repeated until a termination criterion has been satisfied.

The mathematical equation of the model, for the prediction of protein binding, is shown below (Equation 2):

2. $fb=\sum_{j=1}^{k} w2j \left(\frac{(\exp(\sum_{i=1}^{P} xi+wij+bj)-\exp(-\sum_{i=1} xi+wij+bj)}{(\exp(\sum_{i=1}^{P} xi+wij+bj)+\exp(-\sum_{i=1} xi+wij+bj)}+b\right)$

x i (i=1…p) is the input that corresponds to the number of data included in the training of the ANN, I from 1 to 15, wij (i=1…p, j=1…k) weight from input to hidden layer, b j (j=1…k) are biases of the neurons in the hidden layer, k=45 for Wrapper method, w2j (j=1…k) are weights from the hidden to the output layer, b is the bias of the output neuron, and fb is the output.

QSAR-CNN-FFNN

CNN is a powerful machine learning technique. Training a CNN requires a large collection of diverse data, which is not an easy task [42]. However, there is an easy way, we can use a pre-trained CNN, which saves a huge amount of time and effort because fine-tuning a network with transfer learning is usually much faster and easier than training a network from scratch [43]. In this work, transfer learning using the ALEXNET CNN was applied to perform feature extraction and then feature reduction with adjustment in the last six layers by new ones. The CNN model was used to extract representative feature vectors from the penultimate fully connected layer, and then these ”deep features” were used for training an FFNN. The choice of dimension 15 for the feature reduction is to perform a comparison with the features selected by wrapper methods. The details of the 22 layers are reported in Supplemental Material 3. To train the network, holdout cross-validation was used to divide the data randomly 70% for training, 15% for validation, and 15% for testing.

Validation of the models

To determine both the generalizability and the actual predictive capacity of the QSAR models, internal and external validation criteria were used for the validation of the models. The statistical parameters used in our study to check the performance of the models were the coefficient of determination (R^2), the correlation coefficient (R), the predictive squared correlation coefficient (Q^2), and the mean squared error values (MSE) (Equations 3-5):

3. $R^2=1-RSS/SS$
4. $MSE=\sum_i^n=1 \frac{(y_i^{pred}-y_i)^2}{2}$
5. $Q^2=1-PRESS/SS$

The residual sum of squares (RSS) is the difference between the fitted values and the observed values. It refers to the difference between the observation and their mean. The predictive RSS is the difference between the predictions and the observations.

Results

QSAR-Wrapper-FFNN method

The results obtained from the selection of the most important descriptors by the wrapper method, using the correlation coefficient R and the variance inflation factor VIF, showed that 15 descriptors seemed to be the most appropriate. The calculated *VIF*s among the values of the selected descriptors were less than five, indicating that multicollinearity between the selected descriptors is acceptable. Supplemental Material 4 shows the *VIF* values for the selected descriptors and their meanings. To specify the number of hidden neurons required, the procedure detailed above was followed. R (all), MSE (validation), and (validation) criteria were employed for the evaluation of the accuracy of the best model. The best model was chosen according to the minimum MSE (validation) and the maximum R (all), and (validation) [35, 44].

Table 1 shows ten network models developed after the wrapper approach. The results obtained showed that network nine with 45 neurons was the best model with R (all)=0.935, R^2_{train}=0.875, Q^2=0.871, and MSE (validation)=0.015. The best performance of the model had a topology of 15-[45]-1:15 input nodes, one hidden layer with 45 nodes having the hyperbolic tangent as a transfer function, and one output layer with an identity transfer function. The neural networks were implemented using the neural network toolbox for MATLAB [37]. Figure 2 shows a comparison between the experiment and predicted plasma protein binding values for training, validation, and testing sets. The results showed a close correlation between predicted and observed plasma protein binding. The FFNN models were trained with the Levenberg-Marquardt backpropagation training function and gradient descent with momentum weight and bias learning function, and the data were partitioned

Table 1. Selected criteria of the different feed-forward neural networks (FFNNs) obtained by the quantitative structure-activity relationship (QSAR)-Wrapper-FFNN methodology

Number of Hidden Neurons	R (all)	R^2_{train}	Q^2	Mean Squared Error (Validation)
5	0.754	0.539	0.632	0.048
10	0.830	0.679	0.728	0.033
15	0.838	0.694	0.756	0.025
20	0.840	0.724	0.651	0.038
25	0.849	0.712	0.712	0.035
30	0.894	0.810	0.780	0.025
35	0.855	0.707	0.764	0.029
40	0.907	0.814	0.871	0.015
45	**0.935**	**0.875**	**0.871**	**0.015**
50	0.903	0.806	0.821	0.023

Table 2. Selected criteria for the different feed-forward neural networks (FFNNs) obtained from the quantitative structure-activity relationship (QSAR)- convolutional neural network (CNN)-FFNN method

Number of Hidden Neurons	R (all)	R^2_{train}	Q^2	Mean squared error (Validation)
5	0.882	0.812	0.701	0.040
10	0.908	0.832	0.798	0.009
15	0.913	0.856	0.760	0.028
20	0.933	0.857	0.948	0.009
25	0.910	0.834	0.830	0.022
30	0.905	0.830	0.775	0.022
35	0.960	0.905	0.964	0.005
40	0.922	0.832	0.961	0.007
45	0.940	0.874	0.899	0.012
50	0.922	0.830	0.841	0.009

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using holdout cross-validation. The difference between R^2_{train} and Q^2 was equal to 0.004. This difference did not exceed 0.3 indicating the robustness of the model [45].

QSAR-CNN-FFNN method

Regarding the QSAR-CNN-FFNN, the results obtained (Table 2) showed that network seven was the best model with R (all)=0.960, R^2_{train}=0.905, Q^2=0.964, and MSE (validation)=0.005. The best performance of the model had a topology of 15-[35]-1:15 input nodes, one hidden layer with 35 nodes having the hyperbolic tangent as a transfer function, and an identity transfer function in the output layer. Figure 3 shows a comparison between experimental and predicted plasma protein binding for training, validation, and testing sets. The difference between R^2_{train} and Q^2 is equal to 0.059. This difference is less than 0.3 indicating the robustness of the model [45]. For QSAR-CNN-FFNN, the correla-

Table 3. External and internal criteria of the two models

	Parameters	QSAR-Wrapper-FFNN Method	QSAR-CNN-FFNN Method
Internal validation	R (all)	0.935	0.960
	R^2_{train}	0.875	0.905
	Q^2	0.871	0.964
	MSE	0.015	0.005
	MAE	0.090	0.054
	RMSE	0.122	0.069
	$R^2_{adjusted}$	0.874	0.963
External validation	R	0.933	0.982
	Q^2	0.870	0.945
	MSE	0.016	0.007
	MAE	0.094	0.068
	RMSE	0.129	0.085

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Table 4. Comparison with the literature

Method/Ref.	MAE	R^2	R	MSE
Suggested method (QSAR-Wrapper-FFNN)	Train 0.080 Validation 0.090 Test 0.094	Train 0.875 Validation 0.871 Test 0.870	Train 0.935 Validation 0.933 Test 0.933	Train 0.014 Validation 0.015 Test 0.017
Suggested method (QSAR-CNN-FFNN)	Train 0.066 Validation 0.054 Test 0.068	Train 0.905 Validation 0.964 Test 0.945	Train 0.951 Validation 0.972 Test 0.982	Train 0.011 Validation 0.005 Test 0.007
Yuan et al. [27]	Test 0.076			
Sun et al. [25]	Test 0.126			
Kumar et al. [26]				Train 0.869 Test 0.8881
Li et al. [49]		Train 0.86		
Ghafourian et al. [21]	Train 13.25 Validation 14.96	Train 0.717 Validation 0.646	Train 0.681 Validation 0.641	
Moda et al. [50]	-	Test 0.91		

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tion among each pair of our variables was verified and the results are presented in Supplemental Materials 5. In order to obtain an overview of the correlation structure, a heatmap was used to highlight what is important (Figure 4). The question now is how much is too much correlation between our neural network inputs. It seems that the general rule of thumb is that if the simple correlation coefficient between two regressors is greater than 0.8 or 0.9, multicollinearity is a serious problem [46]. However, with all the rules of thumb, we get in the statistics, there is nothing black or white about this. In our case, a prediction with a correlation coefficient of 0.8 does not appear fatal for our regression model. We can say that the above results served our theory, where more descriptors are valuable and welcome because we accept that by removing some of the features we are also discarding some of the data we have about the problem.

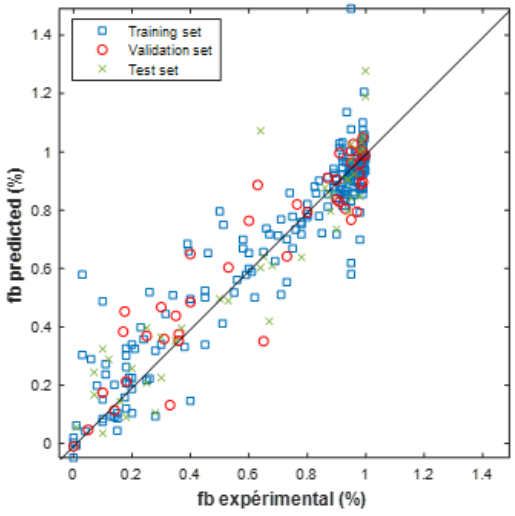
Discussion

Our results described for the first time the use of CNNs as a feature extraction method in QSAR studies for a set of descriptor matrices instead of local pattern extraction from images. One of the huge favorable circumstances of this method is that it does not take much time (only a few minutes) compared to the wrapper method (which took us days and concentration). It also automatically provides us with features that are the best representation of the non-image input, because in this work, we simply constructed the CNN inputs by resizing the 1666 features to 49×34 matrices. We knew that CNNs are used on data that has spatial features, and the fact that it gave us good features to use is that the CNN used discovered that our data has some sort of spatial features. We wanted to know what this deep CNN actually saw, and how it understands the inputs, we feed them. By visualizing the digital values of our matrices, grayscale images were obtained, giving 255 possible different shades of grey

Table 5. Applicability domain of the new method

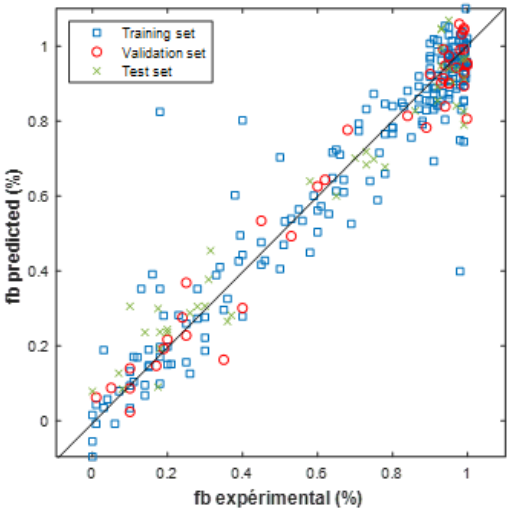
Approaches	Test Inside AD	Test Outside AD
Bounding box	40	2
Leverage	41	1
Euclidean distance (95 percentile)	41	1
Classical KNN (euclidean distance, k=5)	42	0
KNN (euclidean distance (k=25))	42	0

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Figure 2. Comparison between experimental and calculated plasma protein binding predicted by the QSAR-Wrapper-FFNN technique.



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Figure 3. Comparison between experimental and calculated values for plasma protein binding predicted by the QSAR-CNN-FFNN technique.

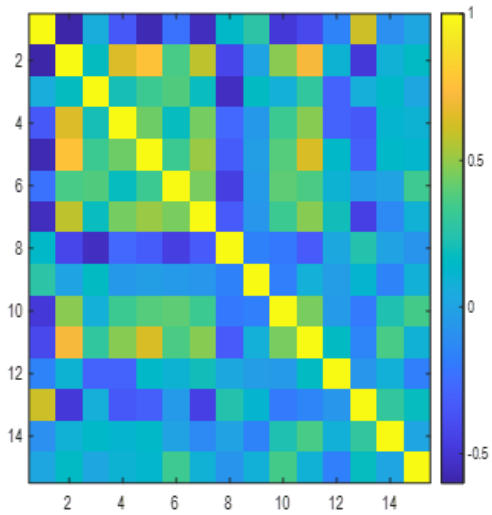
from black to white. Figure 5 shows a sample image of 3-carboxy-4-methyl-5-propyl-2-furanpropionic acid obtained. Numeric values below zero go to zero and those over one go to 255. Values between zero and one are shades of grey, with values between zero and 255. By taking the values of our descriptors and mapping them to the corresponding color suddenly, we can see the structure within our data. We can say that we have made an interface between our data and our brain by visualizing descriptors, in which physical and chemical information about molecules becomes accessible in a new form of fingerprints. They turn into storytellers of molecules secret in just an image.

Comparison of the two models

A statistical evaluation presented in Table 3 was conducted to compare the performance and quality of the predictions by the FFNN of the two models developed for this work. The statistical coefficients of the internal validation for the two models are all acceptable and satisfactory (high R^2_{train}, Q^2 (validation), $R^2_{adjusted}$ and lowest MSE, RMSE, and MAE) and consequently, these models are robust. The quality of the models was also evaluated in terms of external validation criteria. For the wrapper feature extraction method, a value of $Q^2>0.5$ is considered satisfactory and for the QSAR-CNN feature extraction method, the value of $Q^2>0.9$ is considered excellent [45]. We can say that the two models are distinguished by excellent predictive power.

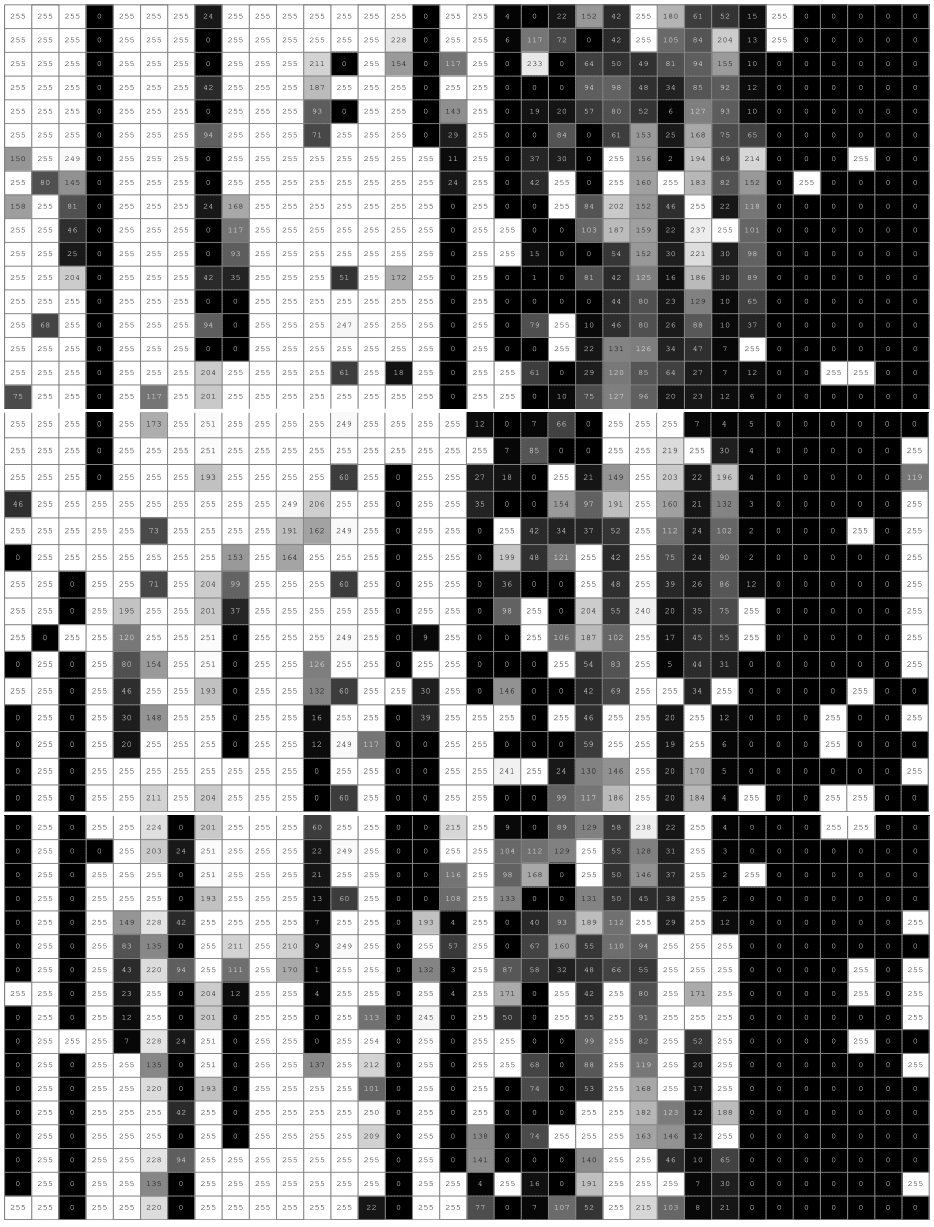
Comparison between models from the literature

A comparison was made between the few models reported in the literature with those developed in this work for the prediction of the binding of drugs to plasma proteins (Table 4). Yawen Yuan et al. constructed a QSAR model for predicting plasma protein binding based on a large training set comprising more than 5000 compounds and proposed a new strategy for constructing models for different binding levels. Lixia Sun et al. de-



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Figure 4. Heatmap of the correlation matrix for the QSAR-CNN-FFNN method.

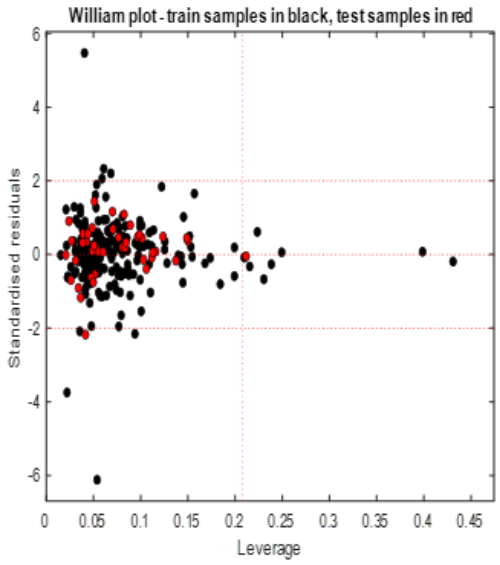


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Figure 5. Image of the input matrices of 3-Carboxy-4-methyl-5-propyl-2-furanpropionic acid with the corresponding numerical values of shade of gray.

veloped QSAR models by six machine learning algorithms with 26 molecular descriptors for predicting the protein-bound fractions of 967 pharmaceuticals. The neural network model yielded an MAE error of 0.126. Rajnish Kumar et al. presented six machine learning algorithms for QSAR models using a database containing 735 drugs. The model with the artificial neural network gave an MSE train=0.869 and MSE val=0.881. Haiyan Li et al. developed a new method called QSAR Plasma Protein Interaction QSAR Analysis (PPI-QSAR) with a database of 65 antibiotics, providing R^2_{train}=0.86 and Q^2=0.72. Taravat Ghafourian et al. collected a database of 794 drugs and used four data mining tools; the best model was boosted trees providing an error MAE train=13.25 and an MAE=14.96. Moda Tiago et al. developed a hologram quantitative structure-activity relationship (HQSAR) on a series of 312 drugs, and they obtained a Q^2=0.72 and R^2=0.91.

We were not interested in evaluating the advantages and disadvantages of these methods because it is quite difficult (each study used different data sets and different modeling approaches) and because the objective of our work was to prove that our new feature selection method



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Figure 6. Plot of the residuals for calculated values of plasma protein binding from the new approach versus their experimental values for training and test sets.

is as effective as these old methods. According to these results, the models developed in this work performed as well as these methods.

Applicability domain

The third principle of the OECD guidelines [47] recommends a defined applicability domain. In this work, the domain of applicability was analyzed with different approaches reported in Table 5. The algorithm and methodology of the proposed approaches are available in the literature [48]. The number of samples inside the applicability domain varied depending on the method used: the bounding box considered two test samples out of the applicability domain, while leverage and Euclidean distance (95 percentile) identified one test sample out of the domain of applicability as shown in Williams plot (Figure 6). Despite its distance from the other observations, our point is close to the regression fitted line since it has a small residual. Classical KNN (Euclidean distance, k=5) and KNN (Euclidean distance (k=25)) showed none of the test samples out of the applicability domain. These results show that we can use the model to predict plasma protein binding for new compounds that have not been tested.

Conclusion

Two different approaches to feature selection in machine learning were tested. These feature selection strategies were followed to produce inputs, which were used in an FFNN to make predictions. Examination of the estimates of external and internal criteria revealed that the QSAR model developed by the new feature selection method is robust, externally predictive, and distinguished by a good applicability domain. The external accuracy of the validation set was calculated by the Q^2 and RMSE, which are equal to 0.945 and 0.085, respectively indicating that the accuracy of NN trained with extracted features from CNNs is slightly better than the accuracy of NN trained with features obtained by the current predictors. This investigation showed that extracting features using CNNs takes less time (only a few minutes) and summarizes much of the information contained in the original features. Contrary to feature selection techniques, we do not have to figure out which features are the best representations of the input, they are learned for us. In the end, according to the OECD principle, we can say that we can use this QSAR-CNN-FFNN model to predict the fraction of plasma protein binding to human plasma for drugs that have not been tested to avoid chemical synthesis and reduce expansive laboratory tests.

Ethical Considerations

Compliance with ethical guidelines

There were no ethical considerations to be considered in this research.

Funding

The authors declare no conflict of interest.

Authors' contributions

Study design: Salah Hanini; Investigation, data collection, and data analysis: Khaouane Affaf; Supervision and Writing–original draft: Samira Ferhat; Writing–review & editing: all authors.

Conflict of interest

The authors declared no conflict of interest.

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